# Preliminary Phytochemical and Antimycobacterial Investigation of Some Selected Medicinal Plants of Endau Rompin, Johor, Malaysia

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Received 30 September 2017; accepted 27 February 2018; available online 1 August 2018

DOI: https://10.30880/jst.2018.10.02.005

**Abstract:** Tuberculosis (TB), the primary cause of morbidity and mortality globally is a great public health challenge especially in developing countries of Africa and Asia. Existing TB treatment involves multiple therapies and requires long duration leading to poor patient compliance. The local people of Kampung Peta, Endau Rompin claimed that local preparations of some plants are used in a TB symptoms treatment. Hence, there is need to validate the claim scientifically. Thus, the present study was designed to investigate the *in vitro* anti-mycobacterial properties and to screen the phytochemicals present in the extracts qualitatively. The medicinal plants were extracted using decoction and successive maceration. The disc diffusion assay was used to evaluate the anti-mycobacterial activity, and the extracts were subjected to qualitative phytochemical screening using standard chemical tests. The findings revealed that at 100 mg/ml concentration, the methanol extract of *Nepenthes ampularia* displayed largest inhibition zone (DIZ=18.67  $\pm$  0.58), followed by ethyl acetate extract of *N. ampularia* (17.67  $\pm$  1.15) and ethyl acetate extract of *Musa gracilis* (17.00  $\pm$  1.00). The phytochemical investigation of these extracts showed the existence of tannins, flavonoids, alkaloids, terpenoids, saponins, and steroids. The pronounced anti-mycobacterial properties displayed by the screened medicinal plants scientifically proved the claim by traditional people of Endau Rompin Johor. It is suggested that the extracts may be considered for further evaluation.

**Keyword:** Antimycobacteria; Phytochemical; Disc diffusion; Endau Rompin.

#### 1. Introduction

Tuberculosis (TB) is a contagious ailment caused by an aerobic pathogenic bacterium called Mycobacterium tuberculosis. This ancient disease is among the world's most deadly epidemics and can happen to anyone, irrespective of sex, age, and nationality [1], [2]. In 2015, World Health Organization (WHO) reported that there were 10.4 million occurrences of TB around the world. About 1.4 million deaths due to TB among HIVnegative persons were documented during 2015. TB accounted for one out of ten causes of mortality globally which more when compared to deadly HIV/AIDS in 2015 [3]. More than half of the global TB cases occur in Asia region (58%), followed by Africa continent (27%). The smaller percentages happen in the East Mediterranean (8%), European region (4%), and America (3%) [4]. Like in any other developing country, TB is still a trait to public health in Malaysia. The trend of TB in the year 2010 in Malaysia

showed a total of 18,517 people have been infected, which is an increase of 6% when compared to the previous year (17,341 cases in the year 2009) [5]. Furthermore, the prevalence of TB was reported as 101 cases per 100 000 population in 2011 [6]. Modern including isoniazid, rifampicin, ethambutol, pyrazinamide, and streptomycin are used in the treated of TB. However, these agents have shortcomings of causing side effects, and the TB-causing bacterium can quickly gain resistance to these drugs [7]. The increase in multi-drug resistant TB and drug-resistant extensively TB strains prevalence in the world is worrisome, and for over 30 years there was no TB medicine introduced into the market [8]. Thus, it is crucial to search for a novel antimycobacterial agent. Due to their chemical variety and important role in the anti-infective agent's development, medicinal plants proffer great hope to overcome these need. For long, plantbased medicines have been used globally in the treatment of different ailments. About 75%

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e-ISSN: 2600-7924/penerbit.uthm.edu.my/ojs/index.php/jst

of the world's populace depend mainly on plant medicines for their primary health care [1]. Jakun is a tribe from a subgroup of Proto-Malay that inhabit Kampung Peta, with the population of around 220 people consisting 67 family units, representing about 2% of the entire Orang Asli in Johor [9]. Jakun tribe living in Kampung Peta depends on their medicinal plants for primary healthcare to treat different diseases traditionally. There was claim by Jakun community in Kampung Peta that 23 species of medicinal plants are used in the treatment of TB and its symptoms [10]. Hence, there need to verify the claim scientifically. Thus, the present study was designed to investigate the in vitro antimycobacterial properties and to screen the phytochemicals present in the extracts qualitatively.

#### 2. Materials and Methods

#### 2.1 Study area

The Endau Romping rainforest, situated border to the north-east of Endau, Johor Darul Takzim and south to Romping, Pahan. The forest (2°25'12.94"N, 103°15'40.94"E) (Fig. 1) is one of those few virgin lowland rainforests remain in southern Peninsular Malaysia. The state government of Johor in 1993 gazetted 870 km<sup>2</sup> of the forest of Taman Negara Johor Endau Rompin (TNJER) as a national park. A village close to the park called Kampung Peta becomes the major entrance to the National Park. Inside the forest lie different species of plant that offer significant sources of shelters, medicines, etc. to the neighboring civilization [10], [11].



**Fig. 1** The location of Kampung Peta in Johor National Park Endau-Rompin

### 2.2 Samples collection and preparation

The plant specimens (Table 1) were collected from Taman Negara Johor Endau Rompin, Johor, Endau Rompin and were identified using the morphometric method in which, the species are recognized based on the characteristic of their leaf, flower shape and/or branching structure. The plant materials were cleaned, chopped into smaller pieces and then dried in a hot oven at 40°C-60°C for 24-72 h [12]–[14]. The dried samples were then ground using a stainless steel blender into a powder which was later sieved. The sieved samples were weight and transferred to a plastic container and stored at room temperature.

Table 1 List of the medicinal plants used in the study

Scientific name	Local name	Family	Part collected	Collected from
Campnosperma	Habong	Anacardiaceae	Shoot	Bununt 49m
auriculatum				02°31.857'N
(Blume) Hook.f.				103°22.624'E
Musa gracilis	Pisang sum	Musaceae	Pseudostem	Sungai Samawak
Holttum				42m 02°31.7491'N
				103°23.807'E
Macaranga	Tudung	Euphorbiaceae	Stem	Pantai Burung 30m
gigantea				02°31.864'N
(Rchb.f				102°24.604'E
&Zoll.)M.A.				
Nepenthes	Sentoyot	Nepenthaceae	Root	Lubuk Bong 43m
ampularia				02°31.542'N
Jack				103°21.706'E

Scaphiur	n Kembang	Sterculiaceae	Stem bark	Ethnobatanical
macropo	dum semangkok			garden 62m
(Miq.) B	eum´			02°31.7491'N
ee ex.He	yne			103°24.845'E

#### 2.3 Extraction

#### 2.3.1 Decoction

The aqueous extract was prepared by immersing the plant materials in a specified volume of distilled water in 1:4 ratios. The extraction mixture was gently heated to the temperature of 60°C in a water bath until the volume of the water was brought down to one-fourth its original volume [15]. Then, the mixture was cooled and strained (filtered) through the Whatman no. 1 filter paper and the filtrate was frozen at -80°C in a freezer and then freeze-dried at -44°C using a freeze drier. The weights of the dried crude extracts were determined.

#### 2.3.2 Successive maceration

The successive maceration method was used for solvent extraction. The powdered plant materials were sequentially macerated with the specified volume of n-hexane, ethyl acetate, and methanol in order of increasing polarity of the solvents in 1:5 ratios in an enclosed flask with occasional shaking. The mixture was kept at room temperature for 24 h. This extraction procedure was repeated three times until complete extraction. The mixture was then strained through a No. 1 Whatman filter paper. The filtrate was later evaporated to a minimum volume a rotary evaporator set at 40°C in a water bath. The weights of the dried crude extracts were determined.

#### 2.4 Phytochemicals investigation

The plant crude extracts were screened for the presence of phytochemicals including flavonoids, tannins, alkaloids, saponins, terpenoids, and steroids using the standard procedures previously described by Bargah (2015), Abdulkadir et al. (2015), and Amabye & Tadesse (2016) [16]–[18].

# 2.5 Test organism and preparation of inoculums

Mycobacterium smegmatis used in the study were obtained from microbiology laboratory of Universiti Tun Hussein Onn Malaysia. The pure isolate was prepared from

the stock culture of *Mycobacterium smegmatis* and then preserved on Middlebrook 7H10 agar medium. The pure culture was then stored at 4°C until further use. From the pure cultures, the inoculum was prepared by subculturing onto Middlebrook 7H9 broth medium. The bacterial suspension density was pre-adjusted to 0.5 McFarland standards.

# 2.6 Determination of Anti-mycobacterial and Antibacterial Activity

Disc diffusion assay previously described by Rafael et al. (2011) with few modifications was used to determine the antimycobacterial activity of the extract. In brief, a filter paper of 5 mm was prepared and sterilized in an autoclave for 15 minutes at 121°C. The filter papers were impregnated with 20 µL of extract at of 12.5, 25, 50 and 100 mg/ml concentrations, and 20 µL of rifampicin (12.5, 25, 50 and 100 µg/mL). The prepared disks were aseptically and carefully transferred onto the Middlebrook 7H10 agar plates inoculated with the cell suspension. The inoculated petri dishes were sealed with parafilm and then incubated at 37°C for 72 hours, after which the diameters of inhibition zone were measured and recorded. This test was performed in triplicate [19].

### 3. Results and Discussions

The inhibitory effect of hexane, ethyl acetate, methanol, and water extracts of C. auriculatum, M. gigantea, M. gracilis N. ampularia, and S. macropodum using agar disk diffusion assay against M. smegmatis at 100, 50, 25, and 12.5 mg/mL concentration are presented in Table 2. It has been shown that at 100 mg/mL concentration, methanolic extract of N. ampularia displayed the largest DIZ (DIZ =  $18.67 \pm 0.58$  mm). On the other hand, at 50 mg/mL, ethyl acetate extract of N. ampularia exhibited the largest DIZ (DIZ =  $13.67 \pm 3.06$  mm). Likewise, ethyl acetate extract of M. gracilis was found to have largest DIZ (DIZ =  $11.33 \pm 0.58$  mm) at the 25 mg/mL concentration. Again, the largest DIZ  $(DIZ = 9.00 \pm 0.00 \text{ mm}) \text{ at } 12.5 \text{ mg/mL was}$ displayed by ethyl acetate extract of M.

gracilis. Interesting, except for hexane extract of N. ampularia, all the hexane extracts were not active in agar disk diffusion assay even at 100 mg/mL concentration. Thus, 13 out of 20 representing 65% of crude extracts were active against the tested bacteria in vitro agar disk diffusion assay. The findings were supported by Fyhrquist et al. (2014) which stated that the diameter of inhibition zone of > 6.00 mm is an indication of antimycobacterial activity against M. smegmatis [20]. The present study showed that methanol and ethyl acetate extracts exhibited the largest diameter of inhibition zone. This is in line with the studies from previous researchers whose proved that ethyl acetate and methanol extracts were active against M. smegmatis [21]-[25]. A literature search showed that the study of the antimycobacterial activity of C. auriculatum, M. gigantea, M. gracilis N. ampularia, and S.

macropodum extracts against M. smegmatis were not previously reported.

The phytochemical screening result found that the extracts contain vast arrays of phytochemicals (Table 3). The phytochemicals including, alkaloids, flavonoids, tannins, saponins, terpenoids, and steroids were found in most of the extracts screened. Arya (2011) stated that wide range of phytochemicals such as were responsible for antimycobacterial flavonoids. activity alkaloids. saponins, terpenoids, and steroids [26]. Solsodomine A, a pyrrole alkaloid isolated Solanum sodomaeum exhibited antimycobacterial activity against intracellulare with 10 µg/mL MIC value [27]. Two flavonoids linaroside and lantanoside were isolated from Lantana camara exhibited antimycobacterial activity against

**Table 2** Diameter of inhibitory zone (DIZ) of plant crude extracts against *M. smegmatis* 

Plant species	Solvent used	100 mg/mL	50 mg/mL	25 mg/mL	12.5 mg/mL	
	Hexane	NA	NA	NA	NA	
C. auriculatum	Ethyl acetate	NA	NA	NA	NA	
	Methanol	$8.67 \pm 0.58$	$7.33 \pm 0.58$	NA	NA	
	Water $10.67 \pm 0.58$ $8 \pm 0.00$		NA	NA		
	Hexane	NA	NA	NA	NA	
M. gigantea	Ethyl acetate	NA	NA	NA	NA	
m. gigamea	Methanol	$14.67 \pm 0.58$	$12.33 \pm 0.58$	11.00 ± 0.00	$8.67 \pm 0.58$	
	Water	$15.33 \pm 0.58$	NA	NA	NA	
	Hexane	NA	NA	NA	NA	
M. gracilis	Ethyl acetate			11.33 ± 0.58	$9.00 \pm 0.00$	
	Methanol	$13.00 \pm 0.00$	$10.33 \pm 0.58$	NA	NA	
	Water	NA	NA	NA	NA	
	Hexane	$9.00 \pm 0.00$	NA	NA	NA	
N. ampularia	Ethyl acetate	$17.67 \pm 1.15$	$13.67 \pm 306$	10.67 ± 3.06	NA	
	Methanol	$18.67 \pm 0.58$	$11.67 \pm 1.53$	$8.00 \pm 1.00$	$7.33 \pm 0.58$	
	Water	$12.00 \pm 1.00$	$10.33 \pm 0.58$	8.333 ± 0.58	$7.33 \pm 0.58$	

	Hexane	NA	NA	NA	NA	
S. macropodum	Ethyl acetate	$12.00 \pm 1.00$	$9.33 \pm 0.58$	$8.00 \pm 1.00$	$7.33 \pm 0.58$	
	Methanol	$9.33 \pm 0.58$	$33 \pm 0.58$ $7.67 \pm 0.58$		NA	
	Water	$10.00 \pm 1.00$	$7.33 \pm 0.58$	$7.00 \pm 0.00$	NA	
RIF (µg/mL)	n/a	$15.67 \pm 0.58$	$10.00 \pm 0.00$	$9.33 \pm 1.15$	$7.33 \pm 0.58$	
DMSO	n/a	NA	NA	NA	NA	

Notes: n/a = not applicable; NA = not active; RIF = Rifampicin; DMSO = Dimethyl sulfoxide. The values were expressed as the mean  $\pm SD$  perform in triplicate

tuberculosis H37Rv by inhibiting 30, 37% of the growth, respectively at 6.25 µg/mL concentration [28]. Tannins, ellagitannin and punicalagin obtained from Combretum molle stem bark were active against M. tuberculosis typus humanus [29]. Triterpenoid, friedelin was isolated from Terminalia avicennioides, which is medicinal plant used by Nupes people of North Central Nigeria in TB treatment. When tested against Bacillus Calmette Guerin (BCG). the compound exhibited antimycobacterial activity with 4.9 µg/mL MIC value [30]. Elsohly et al. (1999) isolated new saponin, jujubogenin 3-*O*-α-Larabinofuranosyl  $(1 \to 2)$ -[3-*O*-(trans)-*p*- coumaroyl-β-D-glucopyranosyl  $(1\rightarrow 3)$ ]- $\alpha$ -Larabinopyranoside from the stems Colubrina retusa. The compound exhibited antimycobacterial activity against intracellulare with the MIC value of 10 µg/mL [31]. From aerial part of Ruprechtia triflora,  $5\alpha$ ,  $8\alpha$ -epidioxyergosta-6, 22-dien-3 $\beta$ -yl stearate which a novel sterol was isolated and exhibited antimycobacterial activity with the MICs of 2-128 µg/mL [32]. Those previous findings indicated that the presence of active compounds was considerably responsible for their antimycobacterial activity.

Table 3 Phytochemical class of crude extracts of selected medicinal plants by chemical analysis

Plant species	Solvents	Alkaloids	Flavonoids	Tannins	Saponins	Terpenoids	Steroids
Campnosperma	Hexane	+	+	+	(-)	+	+
auriculatum	Ethyl acetate	+	+	+	+	+	+
	Methanol	+	+	+	+	+	+
	Water	+	+	(-)	+	(-)	(-)
Musa gracilis	Hexane	+	+	(-)	+	+	+
O	Ethyl acetate	+	+	+	+	+	+
	Methanol	+	+	+	+	+	+
	Water	(-)	+	(-)	+	+	(-)
Macaranga	Hexane	+	(-)	+	+	+	+
gigantea	Ethyl acetate	+	+	+	+	+	+
	Methanol	+	+	+	+	+	+
	Water	+	+	+	+	(-)	+
Nepenthes	Hexane	+	+	+	+	+	+
ampularia	Ethyl acetate	(-)	+	+	+	+	+
conquium to	Methanol	+	+	+	+	+	+
	Water	+	+	+	+	+	(-)
Scaphium	Hexane	+	+	+	(-)	+	+
macropodum	Ethyl acetate	+	+	+	+	(-)	+
	Methanol	+	+	+	+	+	+
	Water	+	+	+	+	+	+

Notes:

<sup>+ :</sup> denotes present; (-) : denotes not present

#### 4. Conclusion

The pronounced antimycobacterial properties displayed by the screened medicinal plants scientifically proved the claim by traditional people of Endau Rompin Johor. It is suggested that the extracts may be refined and standardized to be used as an alternative or complementary medicine and further studies should be carried out to isolate the bioactive compounds which could be potential anti-TB drug leads.

# Acknowledgement

The authors would like to thanks the Office for Research, Innovation, Commercialization and Consultancy Management, Universiti Tun Hussein Onn Malaysia (UTHM) for assisting with the research grant (UTHM Grant Contract, Vot No: U555) that supported the study.

# References

- [1] Sanusi, S.B., Bakar, M.F.A, Mohamed, M., Sabran, S.F. and Mainasara, M.M. (2017). "Southeast Asian Medicinal Plants as a Potential Source of Antituberculosis Agent" Evidence-Based Complement. *Altern. Med.*, vol. 2017 Article ID 7185649, 39 pages.
- [2] Bueno-Sánchez, J.G., Martínez-Morales, J.R., Stashenko, E.E. and Ribón, W. (2009). "Anti-tubercular Activity of Eleven Aromatic and Medicinal Plants Occurring in Colombia" *Biomedica*, vol. 29. no. 1 pp. 51–60.
- [3] Almatar, M., Almandeal, H., Kayar, B. and Köksal, F. (2017). "New Drugs for the Treatment of Mycobacterium Tuberculosis Infection" *Biomed. Pharmacother.*, vol. 91. pp. 546–558.
- [4] Radji, M. Kurniati, M. and Kiranasari, A. (2015). "Comparative Antimycobacterial Activity of Some Indonesian Medicinal Plants Against Multi-Drug Resistant Mycobacterium Tuberculosis" *J. Appl. Pharm. Sci.*, vol. 5. no. 1 pp. 19–22.
- [5] Sanusi, S.B., Talip, B.A. and Mohamed, M. (2017). "The Descriptive Study of Knowledge and Awareness of Tuberculosis Among Students in

- Universiti Tun Hussein Onn Malaysia" *J. Sci. Technol.*, vol. 9. no. 1 pp. 15–19.
- [6] Margolis, B., Al-Darraji, H.A.A., Wickersham, J.A., Kamarulzaman, A. and Altice, F.L. (2013). "Prevalence of Tuberculosis Symptoms and Latent Tuberculous Infection Among Prisoners in Northeastern Malaysia" *Int. J. Tuberc. Lung Dis.*, vol. 17. no. 12 pp. 1538–1544.
- [7] Adaikkappan, P., Kannapiran, M. and Anthonisamy, A. (2012). "Anti-Mycobacterial Activity of Withania Somnifera and Pueraria Tuberosa against Mycobacterium Tuberculosis H 37 Rv" *J. Acad. Indus. Res.*, vol. 1. pp. 153–156.
- [8] Gautam, R., Saklani, A. and Jachak, S.M. (2007) "Indian Medicinal Plants as a Source of Antimycobacterial Agents" *J. Ethnopharmacol.*, vol. 110. no. 2 pp. 200–234.
- [9] Mainasara, M.M., Bakar, F.M.A., Mohamed, M., Alona, C. and Sanusi, S.B. (2017). "Ethnomedical Knowledge of Plants Used for the Treatment of Breast Cancer by Jakun community in Kampung Peta Endau Rompin Johor, Malaysia" *J. Sci. Technol.*, vol. 9. no. 1 pp. 42–49.
- [10] Sabran, S.F., Mohamed, M. and Bakar. M.F.A. (2016). "Ethnomedical Knowledge of Plants Used for the Treatment of Tuberculosis in Johor, Malaysia" Evidence-Based Complement. Altern. Med., vol. 2016, (2016). Article ID 285045, 12 pages.
- [11] Ismaila, I., Linatoca, A.C., Mohamed, M. and Tokiman, L. (2015). "Documentation of Medicinal Plants Traditionally used by the Jakun People of Endau-Rompin (PETA) for Treatment of Malaria-Like Symptom" *J. Teknol.*, vol. 77. no. 31 pp. 63–69.
- [12] Derbie, A., Young, A. and Keum, S. (2016). "Effect of Extraction Solvent and Various Drying Methods on Polyphenol Content and Antioxidant Activities of Yuzu (Citrus junos Sieb ex Tanaka)" *J. Food Meas. Charact.*, vol. 11. no. 2 pp. 576–585.
- [13] Ghasemi, A., Salehi, S. and Craker, L. (2017). "Effect of Drying Methods on Qualitative and Quantitative Properties of Essential Oil from the Aerial Parts of

- coriander," *J. Appl. Res. Med. Aromat. Plants*, vol. 4. pp. 35–40.
- [14] Vongsak, B., Sithisarn, P., Mangmool, S., Thongpraditchote, S., Wongkrajang, Y. and Gritsanapan, W. (2013). "Maximizing Total Phenolics, Total Flavonoids Contents and Antioxidant Activity of Moringa Oleifera Leaf Extract by the Appropriate Extraction Method" *Ind. Crops Prod.*, vol. 44. pp. 566–571.
- [15] Handa, S.S., Khanuja, S.P.S., Longo, G. and Rakesh, D.D. (2008). Extraction Technologies for Medicinal and Aromatic Plants, 1stedn ed. Italy: United Nations Industrial Development Organization and the International Centre for Science and High Technology.
- [16] Bargah, R. (2015). "Preliminary Test of Phytochemical Screening of Crude Ethanolic and Aqueous Extract of Moringa pterygosperma Gaertn" *J. Pharmacogn. Phytochem.*, vol. 4. no. 1 pp. 7–9.
- [17] Abdulkadir, S.I., Nasir, I.A., Sofowora, A., Yahaya, F., Ahmad, A.A. and Hassan, I.A. (2015). "Phytochemical Screening and Antimicrobial Activities of Ethanolic Extracts of Moringa oleifera Lam on Isolates of Some Pathogens" *J. Appl. Pharm.*, vol. 7. no 4.
- [18] Amabye T.G. and Tadesse, F.M. (2016). "Phytochemical and Antibacterial Activity of Moringa Oleifera Available in the Market of Mekelle" *J. Anal. Pharm. Res.*, vol. 2. no. 1 pp. 1–4.
- [19] Rafael, J., Peixoto, O., Silva, G.C., Costa, R.A., Fontenelle, L.D.S., Hitzschky, G., Vieira, F., Adauto, A., Filho, F., Helena, R. and Vieira, F. (2011). "In Vitro Antibacterial Effect of Aqueous and Ethanolic Moringa Leaf Extracts" *Asian Pac. J. Trop. Med.*, vol. 4. no. 3 pp. 201–204.
- [20] Fyhrquist, P., Laakso, I., Garcia Marco, S., Julkunen-Tiitto, R. and Hiltunen, R. (2014). "Antimycobacterial Activity of Ellagitannin and Ellagic Acid Derivate Rich Crude Extracts and Fractions of Five Selected Species of Terminalia used for Treatment of Infectious Diseases in African Traditional

- Medicine" *South African J. Bot.*, vol. 90. pp. 1–16.
- [21] Syahputra, G., Sari, M. and Wien, K. (2016). "Anti-mycobacterial Activity of Methanol Plants Extract to against Mycobacterium Bovis and Mycobacterium smegmatis" *Pros. Semin. Nas. Masy. Biodiversitas Indones.*, vol. 2. no. 2 pp. 1882–1887.
- [22] Newton, S.M., Lau, C., Gurcha, S.S., Besra, G.S. and Wright, C.W. (2002). "The Evaluation of Forty-three Plant Species for in Vitro Antimycobacterial Activities; Isolation of Active Constituents from Psoralea Corylifolia and Sanguinaria Canadensis" *J. Ethnopharmacol.*, vol. 79. no. 1 pp. 57–67.
- [23] Tamreihao, K. (2017). "In-vitro Antimycobacterial Activities of Endophytic Bacteria Associated with Medicinal Plant of Manipur" *J. Bacteriol. Mycol.*, vol. 4. no. 4 pp. 1-4.
- Boligon, A.A., Agertt, V., Janovik, V., [24] C., Ritiel. Campos, M.M.A.. Guillaume, D., Athayde, L. and Santos, A.R.S. (2012)."Antimycobacterial Activity of the Fractions Compounds from Scutia Buxifolia" Brazilian J. Pharmacogn., vol. 22. no. 1 pp. 45–52.
- [25] Bhunu, B., Mautsa, R. and Mukanganyama, S. (2017). "Inhibition of Biofilm Formation in Mycobacterium Smegmatis by Parinari Curatellifolia Leaf Extracts" *BMC Complement. Altern. Med.*, vol. 17. no. 1 p. 285.
- [26] Arya, V. (2011). "A Review on Anti-Tubercular Plants" *Int. J. PharmTech Res.*, vol. 3. no. 2 pp. 872–880.
- [27] Kishore, N., Mishra, B.B., Tripathi, V. and Tiwari, V.K. (2009) "Alkaloids as Potential Anti-Tubercular Agents" *Fitoterapia*, vol. 80. no. 3 pp. 149–163.
- [28] Begum, S., Wahab, A. and Siddiqui, B.S. (2008). "Antimycobacterial Activity of Flavonoids from Lantana Camara Linn" *Nat. Prod. Res.*, vol. 22. no. 6 pp. 467–470.
- [29] Thiel, W., Asres, K., Bucar, F., Edelsbrunner, S., Kartnig, T. and Ho, G. (2001). "Investigations on Antimycobacterial Activity of Some Ethiopian Medicinal Plants" *Phyther*.

- Res., vol. 15. no. 4 pp. 323–326.
- [30] Mann, A., Ibrahim, K., Oyewale, A.O., Amupitan, J.O., Fatope, M.O. and Okogun, J.I. (2011). "Antimycobacterial Friedelaneterpenoid from the Root Bark of Terminalia avicennioides" *Am. J. Chem.*, vol. 1. no. 2 pp. 52–55.
- [31] Elsohly, H.N., Danner, S., Li, X.C., Nimrod, A.C. and Clark, A.M. (1999). "New Antimycobacterial Saponin from Colubrina retusa" *J. Nat. Prod.*, vol. 62. no. 9 pp. 1341–1342.
- [32] Woldemichael, G.M., Franzblau, S.G., Zhang, F., Wang, Y. and Timmermann, B.N. (2003). "Inhibitory Effect of Sterols from Ruprechtia Triflora and Diterpenes from Calceolaria Pinnifolia on the Growth of Mycobacterium Tuberculosis" *Planta Med.*, vol. 69. no. 7 pp. 628–31.